# **New Journal of Physics**

The open access journal at the forefront of physics

Deutsche Physikalische Gesellschaft DPG

Published in partnership with: Deutsche Physikalische Gesellschaft and the Institute of Physics

## PAPER

CrossMark

**OPEN ACCESS** 

RECEIVED 12 April 2019

REVISED

16 July 2019

24 July 2019

9 August 2019

PUBLISHED

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# Theory of active particle penetration through a planar elastic membrane

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Keywords: membranes, elasticity, active particles

Supplementary material for this article is available online

# Abstract

With the rapid advent of biomedical and biotechnological innovations, a deep understanding of the nature of interaction between nanomaterials and cell membranes, tissues, and organs, has become increasingly important. Active penetration of nanoparticles through cell membranes is a fascinating phenomenon that may have important implications in various biomedical and clinical applications. Using a fully analytical theory supplemented by particle-based computer simulations, the penetration process of an active particle through a planar two-dimensional elastic membrane is studied. The membrane is modeled as a selfassembled sheet of particles, uniformly arranged on a square lattice. A coarse-grained model is introduced to describe the mutual interactions between the membrane particles. The active penetrating particle is assumed to interact sterically with the membrane particles. State diagrams are presented to fully characterize the system behavior as functions of the relevant control parameters governing the transition between different dynamical states. Three distinct scenarios are identified. These compromise trapping of the active particle, penetration through the membrane with subsequent self-healing, in addition to penetration with permanent disruption of the membrane. The latter scenario may be accompanied by a partial fragmentation of the membrane into bunches of isolated or clustered particles and creation of a hole of a size exceeding the interaction range of the membrane components. It is further demonstrated that the capability of penetration is strongly influenced by the size of the approaching particle relative to that of the membrane particles. Accordingly, active particles with larger size are more likely to remain trapped at the membrane for the same propulsion speed. Such behavior is in line with experimental observations. Our analytical theory is based on a combination of a perturbative expansion technique and a discrete-tocontinuum formulation. It well describes the system behavior in the small-deformation regime. Particularly, the theory allows to determine the membrane displacement of the particles in the trapping state. Our approach might be helpful for the prediction of the transition threshold between the trapping and penetration in real-space experiments involving motile swimming bacteria or artificial active particles.

# 1. Introduction

As one of the most fundamental components in biological systems, the cell membrane defines and protects the cell and is selectively permeable for ions and organic molecules, allowing to control the movement of required chemicals into the cell and of unwanted products out of the cell. It is now possible not only to reassemble cell membranes artificially [1], but also to design synthetic membranes with properties tailored to the needs of 21st centuries societies [2–4]. In fact, synthetic membranes are now routinely used already for applications from water purification [5, 6] to dialysis [7, 8] and can be regarded as a paradigmatic success of biomimetics [9–11]. Future perspectives for the usage of synthetic membranes involve problems like targeted gene and drug delivery to (cancer) cells [12–24] or, more

generally, the delivery of cargo to the interior of synthetic droplets, requiring a precise understanding of the interaction of motile particles with synthetic and biological membranes. Evidence from previous studies has shown that the physical uptake by living cells is strongly affected by the particle and membrane physicochemical and functional properties [25–31].

While membranes comprising active inclusions, e.g. in the form of embedded proteins creating a stress on the membrane, have been studied for decades [32–34], the penetration of active particles through the membrane is less explored [14, 35] with the few existing studies focusing on nano- and biotechnology perspectives. In particular, penetration of nanoparticles through a membrane has been studied using dissipative particle dynamics simulations, focusing on effects of particle shape [14] and surface-structure [36]. In addition, molecular dynamics simulations have been employed to investigate the penetration of fullerens through lipid membranes [37]. Recent studies have also explored interactions of active particles with membranes, from a more physical point of view, but did not focus on particle penetration [38–40]. For a 1D membrane, we have recently performed a corresponding investigation [41].

Conversely to most of the above works, here we explore the penetration of an active particle through a 2D synthetic membrane from a physics perspective, aiming at predicting overall properties such as the membrane shape or the parameter domain leading to penetration starting from coarse microscopic details. We focus on a minimal model membrane that can be realized in principle using as building blocks microparticles interacting via elastic forces [42–48]. Other types of interactions, such as dipolar interactions may be considered as well to model self-assembled chains and sheets [49–61]. To predict the state diagram, informing us about the parameter domains where particles can penetrate through the membrane and where they cannot, we systematically derive a continuum description of the membrane. We compare our results with particle-based computer simulations, finding close quantitative agreement regarding the transition between trapping and penetrating states, membrane shape and dynamics. Our analytical closed-form expressions might help to predict the properties of synthetic membranes, e.g. regarding the speed and size of particles which will be able to penetrate through them.

Below, we first define our model (section 2), followed by a brief discussion of the relevant parameters and of the 2D membrane dynamics as induced by the active particle approaching it (section 3). Here, besides a trapping state in which the membrane is deformed in the final state and does not allow the particle to pass, we find two scenarios of penetration. The first of these corresponds to the particle breaking through the membrane, followed by a complete self-healing of the membrane, which might be the desired behavior when delivering cargo towards a synthetic droplet or a healthy cell. The second scenario of penetration occurs mainly for larger particles, creating a hole in the membrane with a size exceeding the interaction range of the membrane components. This situation is accompanied by a partial fragmentation of the membrane structure into isolated particles. Following this qualitative discussion, we systematically explore the corresponding state diagram using numerical simulations, showing for which parameter combinations which of these three states prevails, and we develop a detailed analytical theory (section 4). The latter is able to predict essentially the entire state diagram as well as the shape and the dynamics of the membrane, in close quantitative agreement with our simulations. Interestingly, the transition between self-healing and non-healing states is sharp, suggesting that there is a critical size for particles that pass a membrane by causing significant damage. This also suggests that if one were to permanently damage the membrane in our minimal model (and perhaps similarly in practice to treat cancer cells), one needs to use particles with a certain minimal size. Finally, concluding remarks summarizing our findings are contained in section 5.

# 2. System setup

We examine the penetration mechanism of a non-fluctuating membrane by an active particle moving under the action of a constant propulsion force  $F_0$ . We assume that the persistence length of the self-propelling active particle is larger than the distance initially separating the particle from the membrane. Correspondingly, we focus on the limiting case of vanishing rotational diffusion. This implies that the particle essentially moves along a straight trajectory without changing its orientation [62–73]. The active particle may represent a swimming microorganism [74–79] or an artificial microrobot that can be manipulated by controlled external fields [80–83].

In our model, the membrane is composed of *N* identical spherical beads (or vertices) of radius *a*, uniformly arranged on a square lattice of size  $L \times L$ , rotated by 45° around the *z* axis, the latter directed normal to the membrane, as schematically illustrated in figure 1. We denote by *h* the lattice spacing after initialization. The membrane is immersed in a Newtonian fluid, characterized by a constant dynamic viscosity  $\eta$ . We support the membrane at its periphery (the particle displacements are zero for  $x, y = \pm L/2$ ) and assume periodic boundary conditions in the transverse directions (*x*, *y*). Moreover, we suppose that the mutual interactions between the membrane particles are pairwise additive and described by forces that depend only on the difference of



**Figure 1.** Graphical illustration of the system setup. (a) An active particle of radius *R* moving through an effective driving force  $F_0$  toward a two-dimensional membrane composed of *N* particles of radius *a*. The membrane particles are initially arranged on a square lattice of dimension  $L \times L$  and spacing *h*, rotated by 45° around the *z* axis, the latter oriented normal to the plane of the membrane. The membrane is centered about the origin and clamped at its periphery. Periodic boundary conditions are imposed in both *x* and *y* directions. We assume that the membrane particles are subject to steric and elastic pairwise interactions. The system is fully immersed in a Newtonian viscous fluid of shear viscosity  $\eta$ . (b) Schematic illustration of the lattice artucture composing the model membrane. For future reference, the eight nearest neighbors of the particle at the center of the lattice are identified by numbers (1–8). Elastic springs are also inserted along the lattice diagonals but are not displayed here for reasons of clarity.

coordinates of each two neighboring particles. Representing the membrane as a collection of spherical beads arranged on vertices has extensively been employed as a coarse-grained model for cell membranes, see, for instance [42–48].

Typically, various types of interactions may occur among membrane particles including steric and elastic interactions. For instance, steric interactions can be imposed by membrane phospholipids chains and other biomolecules [84–86], whereas intermolecular coupling between the lipid bilayer and the cytoskeleton network gives rise to elastic interactions [87–90]. Accordingly, the total potential energy of the membrane here is written as a sum of two distinct contributions as

$$\mathcal{U} = 4\epsilon \sum_{\substack{i,j=1\\j < i}}^{N} N_{ij} \left( \left( \frac{\sigma}{r_{ij}} \right)^6 \left( \left( \frac{\sigma}{r_{ij}} \right)^6 - 1 \right) + \frac{1}{4} \right) + \frac{k}{2} \sum_{i=1}^{N} \sum_{\substack{j \in \mathcal{N}(i)\\j < i}} (r_{ij} - \xi r_{0ij})^2, \tag{1}$$

wherein  $r_{ij} = |r_{ij}|$  is the distance between particles *i* and *j*, and  $r_{ij} = r_i - r_j$ . In addition,  $\epsilon$  is an energy scale associated with the Weeks–Chandler–Andersen (WCA) pair-potential [91],  $\sigma = 2a$  is the diameter of the particles,  $N_{ij} = H(r_{\rm C} - r_{ij})$ , with  $H(\cdot)$  denoting the Heaviside step function, and  $r_{\rm C} = 2^{1/6}\sigma$  is a finite cutoff distance beyond which the steric interactions energy vanishes. Furthermore, *k* is the elastic constant of the harmonic springs coupling each particle to its four nearest and four next-nearest neighbors,  $r_{0ij}$  is the rest length of the springs, and  $\xi \in (0, 1]$  is a prestress parameter. Here, we use the notation  $\mathcal{N}(i)$  to denote the set of nearest and next-nearest neighbors of the *i*th membrane particle. For real cell membranes, the lattice spacing *h* may, for instance, be viewed as an average distance between cytoskeleton-bilayer connection sites. In addition, the elastic constant *k* may be connected to the shear modulus of the cytoskeleton network, the order of magnitude of which is about  $10^{-6}$  N m<sup>-1</sup>.

For the sake of simplicity, we neglect all possible hydrodynamic interactions between particles. Moreover, we assume that the particles are small enough or sufficiently matched in density to the surrounding fluid for the

influence of gravity to be neglected, and large enough for the effect of thermal fluctuations to be neglected. In addition, we assume throughout this work that the size of the active particle is comparable to or larger than that of the membrane particles.

The corresponding interaction force acting on the *i*th membrane particle is obtained by differentiating the potential energy described by equation (1) with respect to the particle position [92] as  $F_i = -\partial U/\partial r_i$ . Accordingly,

$$F_{i} = 48\epsilon \sum_{\substack{j=1\\j\neq i}}^{N} \frac{N_{ij}}{r_{ij}} \left(\frac{\sigma}{r_{ij}}\right)^{6} \left(\left(\frac{\sigma}{r_{ij}}\right)^{6} - \frac{1}{2}\right) \hat{r}_{ij} + k \sum_{j \in \mathcal{N}(i)} (\xi r_{0ij} - r_{ij}) \hat{r}_{ij},$$
(2)

where  $\hat{r}_{ij} = r_{ij}/r_{ij}$  is a unit distance vector.

At small length scales, aqueous systems are characterized by small Reynolds numbers, so that viscous forces dominate over inertial forces. The resulting overdamped dynamics can therefore be adequately described within the framework of linear hydrodynamics [93, 94]. Accordingly, the translational velocity of the membrane particles *V<sub>i</sub>* is linearly coupled to the forces acting on their surfaces via the hydrodynamic mobility functions [95–98]. The latter are second-order tensors, which simply reduce to scalar quantities when considering motion in an unbounded medium and neglecting the fluid-mediated hydrodynamic interactions between the particles. Specifically,

$$V_i = \frac{\mathrm{d}r_i}{\mathrm{d}t} = \mu(F_i + F_i^{\mathrm{ext}}),\tag{3}$$

where  $\mu$  denotes the translational self-mobility functions of the membrane particles. This is given by the usual Stokes formula for an isolated sphere in an infinite fluid domain as  $\mu = 1/(6\pi\eta a)$ . In addition,  $F_i^{\text{ext}}$  represent the external force exerted by the active particle due to the steric interactions with the membrane particles. These pair interactions are modeled via a soft repulsive WCA potential as in equation (1) for which  $\sigma = R + a$ , with R denoting the radius of the active particle.

We introduce at this point an additional cutoff length  $\ell$  beyond which the elastic interactions are set to zero. Accordingly, the elastic potentials are also shifted to this cutoff length, so as to ensure that the resulting potentials are continuous.

# 3. Trapping, penetration, and self-healing

Having introduced a model for our membrane and derived the corresponding equations governing the translational dynamics of the particles composing the membrane, we next study in detail the dynamical states emerging from the interaction between an active particle propelling toward the membrane. For that purpose, we solve numerically the set of ordinary differential equations in time given by equations (2) and (3) using a standard Euler scheme with adaptive time stepping [99]. Before the active particle starts to interact with the membrane particles, we assume that the lattice spacing *h* is identical to the cutoff length scale  $r_{\rm C}$  associated with the WCA pair potential. In addition, we assume that the rest length of the elastic springs is equal to the initial interparticle separation, i.e.  $r_{0ij} = h$  for the pairs of particles located along the lattice axes, and  $r_{0ij} = \sqrt{2} h$  for the pairs along the diagonal. Under these conditions, the membrane is initially at equilibrium, on account of the periodic boundary conditions imposed along the transverse directions (*x*, *y*). We further mention that requiring  $h = r_{\rm C}$  is equivalent to considering a constant ratio  $h/a = 2^{7/6}$ . Unless stated otherwise, we consider throughout the present article a membrane composed of N = 450 particles and set the prestress parameter as  $\xi = 0.9$ .

We now introduce the reduced activity

$$E = \frac{aF_0}{\epsilon},\tag{4}$$

which represents a balance between the magnitude of the active driving force  $F_0$  and the steric forces at particle contact. Further, we define the reduced stiffness

$$\kappa = \frac{akh}{2\epsilon},\tag{5}$$

which quantifies the importance of the elastic forces relative to the steric forces. The prefactor one half follows from theoretical considerations as will be shown in the sequel. In addition, we introduce the size ratio

$$\delta = \frac{R}{a} \tag{6}$$

 Table 1. Dimensionless numbers characterizing the system in the trapping and penetrating states.

Dimensionless number	Expression	Denomination
$ \frac{E}{\kappa} \\ \delta \\ \lambda \\ P_0 $	$\frac{\frac{aF_0}{\epsilon}}{\frac{akh}{2\epsilon}}$ $\frac{\frac{R}{a}}{\frac{\ell'}{h}}$ $\frac{E}{\kappa} = \frac{2F_0}{kh}$	Reduced activity Reduced stiffness Size ratio Scaled cutoff distance Admittance

to denote the radius of the active particle relative to that of the membrane particles. Finally, we define

$$\lambda = \frac{\ell}{h} \tag{7}$$

as a scaled cutoff distance beyond which the elastic interactions vanish. Unless otherwise stated, we set  $\lambda = 3/2 > \sqrt{2}$ , such that the pair-interactions between the membrane particles are restricted to the four nearest and four next-nearest neighbors only.

An additional dimensionless parameter that we denominate as 'admittance', is introduced to quantify the penetration capability of the active particle. It is defined based on the above definitions of E and  $\kappa$  and expresses the ratio between active and elastic forces. Specifically,

$$P_0 = \frac{E}{\kappa} = \frac{2F_0}{kh}.$$
(8)

Here, the admittance serves to quantify a criterion of whether or not the active particle passes through the membrane. For ease of reference, the explicit expressions of the key dimensionless numbers characterizing the states of the system are listed in table 1.

To get a first intuition of the possible membrane dynamics, we display the different observed scenarios in figure 2. For low admittance ( $P_0 = 1$ , top row and movie S1 available online at stacks.iop.org/NJP/21/083014/ mmedia in the Supporting Information), the membrane starts to deform when the motile active particle comes close, but only up to some point, reaching a steady state of constant membrane shape and fixed position of the active particle (see figure 2, panels (c) and (d)). When increasing the admittance to  $P_0 = \sqrt{10}$  (second row and movie S2), the membrane no longer reaches a steady state, but the active particle breaks through the membrane, leaving a hole that starts to self-heal once the particle has left the membrane particles behind. Here, the membrane evolves back towards its original configuration, as it would be desired, e.g. when delivering cargo to the inside of a healthy cell, the membrane of which we would want to remain intact. The membrane dynamics qualitatively changes when using larger particles instead ( $\delta = 7$ ) and strongly enhancing the admittance to  $P_0 = 100\sqrt{10}$  (third row and movie S3). In this situation, the particle breaks through the membrane and creates a permanent hole. The four particles located around the center of the membrane in figure 2(l) remain isolated because the range of the internal membrane interactions is shorter than the separation distance of these four particles from the rest of the membrane. Such a behavior is even more pronounced for significantly larger particles ( $\delta = 13$ ) (bottom row and movie S4) where the membrane is partially fragmented into four clusters of particle triplets and four clusters of particle sextuplets (figure 2(p)), after the active particle has penetrated through the membrane.

In figure 3, we present a state diagram indicating the system behavior in the parameter space ( $\kappa$ , E). As already mentioned, the membrane is composed of N = 450 particles. Here, we set  $\delta = 1$ . Depending on the ratio between the control parameters  $\kappa$  and E, we observe that the active particle either passes through the membrane to reach the other side (red triangles) or remains trapped (blue rectangles). The transition between the two states can be described by a linear hypothesis of the form  $P_0 = 1$ . Accordingly, penetration events occur when the membrane restoring forces consisting of elastic contributions become weaker than the damaging force resulting from the steric interactions with the active particle. After full penetration has occurred, the membrane self-heals and relaxes back to its initial equilibrium configuration.

In figure 4, we show dynamical state diagrams in the planes of the control parameters (a) ( $\delta$ , E) for  $\lambda = 3/2$ , and (b) ( $\delta$ ,  $\lambda$ ) for E = 1. To limit the parameter space, we set in both diagrams the reduced stiffness to  $\kappa = 10^{-2}$ . We observe that the transition between the trapping and penetration states can also be enabled by varying the size ratio  $\delta$  (figure 4(a)). Accordingly, the penetration capability through a membrane is not only determined by the system admittance, but also by the size of the active particle relative to that of the membrane particles. This is in agreement with earlier experimental investigations indicating that particle size may strongly affect the uptake efficiency and kinetics [100–104]. Consequently, an active particle with a size larger than that of the membrane



**Figure 2.** Snapshots of particle-based computer simulations illustrating the membrane conformation in the trapping and penetration states at different time intervals scaled by the unit simulation time  $t_{\rm S} = \eta L^3/\epsilon$ . At time t = 0, the active particle begins interacting with the membrane particles (cyan circles). Here, we set the reduced stiffness to  $\kappa = 10^{-2}$  in all these simulations. For clarity, the membrane particles located in the planes x = 0 and y = 0 within the interaction range of the elastic potentials are shown as blue disks linked by dashed lines. The first row (panels (a)–(d)) displays the membrane dynamics in the trapping state, for a size ratio  $\delta = 1$  and a reduced activity  $E = 10^{-2}$ . Since the driving force is not strong enough compared to the membrane restoring forces, the active particle remains trapped near the membrane. The second row (panels (e)–(h)) represents the time frames during penetration with subsequent self-healing, for  $\delta = 1$  and  $E = 10^{-1.5}$ . In this state, the membrane recovers its initial planar shape after the active particle has passed through it. Next, the third row (panels (i)–(l)) contains the frame series during penetration without self-healing, for  $\delta = 7$  and  $E = \sqrt{10}$ . The membrane remains permanently damaged after penetration as the mutual distance between the four depicted fragmented particles becomes larger than the scaled cutoff distance  $\lambda$ . The bottom row (panels (m)–(p)) further illustrates the penetration stet without self-healing, for  $\delta = 13$  and  $E = \sqrt{10}$ . Due to the relatively large size of the active particle, the membrane is partially fragmented around its center into four clusters of particle triplets and four clusters of particle setuplets, the distance between these particles and the remainder of the membrane being larger than the cutoff length  $\ell$ . The red disks represent the positions of the active particle, which are out of the field of view in some panels. Because of the pronounced difference between the scales along t

particles is more likely to remain trapped. It is worth noting that, in the considered range of parameters, the transition has been found to only depend on the admittance  $P_0$  for our simplistic 1D model membrane studied in a previous work [41]. For large values of the size ratio and small scaled cutoff distance  $\lambda$ , the penetration process may also occur without subsequent self-healing of the membrane. This situation is accompanied by partial fragmentation of the membrane, during which a number of particles around the center remain isolated from the remainder of the membrane, creating a permanent hole in the membrane. The number of fragments largely depends on the propulsion speed and the size ratio. Interestingly, the membrane was also observed to become partially fragmented into clusters of four quadruplets for ( $\delta$ , E) = (9, 1) or a combination of four triplets and four sextuplets of membrane particles for ( $\delta$ , E) = (13,  $\sqrt{10}$ ). This effect points to an interesting size effect of the membrane. For large scaled cutoff distances (figure 4(b)), the penetration capability decreases, yet the transition between the trapping and penetration states is weakly dependent on the scaled cutoff distance  $\lambda$ . In these situations, the membrane self-heals after particle penetration because the displaced membrane particles remain within the interaction range of the restoring elastic forces.

Having investigated the penetration mechanism of a single active particle self-propelling toward the membrane it is worth commenting on the collective penetration of a large number of active particles. Depending on the density and activity of the penetrating particles as well as on the physical properties of the membrane, the penetration of a group of active particles may show a behavior different from the one observed for a single particle. In order to probe this effect in some detail, we consider  $N_{\rm P}$  active particles initially arranged on a square





lattice around the center of the membrane. The orientation of this square lattice relative to that of the membrane is determined by the number of active particles. The active particles move under the action of equal propulsion forces and are initialized at the same vertical distance to the membrane. The initial Cartesian coordinates of the active particles are listed in table 2 and illustrated for clarity in figure 5.

In figure 6, we present a state diagram in the parameter space  $(\kappa, N_P)$  for  $\delta = 1, \lambda = 3/2$ , and E = 1. We quantify the proportion of the active particles that pass through the membrane by the ratio *r*. Here, r = 0 corresponds to the situation in which all the active particle are fully trapped while r = 1 corresponds to the penetration of all the particles. The latter scenario is always accompanied by a subsequent self-healing considering the present set of parameters. We observe that, as the number of active particles gets larger, the penetration capability increases. This behavior is justified by the fact that, as the number of active particles increases, the forces damaging the membrane become larger than the elastic forces. Consequently, more active particles together are able to break through the membrane even if a single particle would be trapped. In this context, Kaiser *et al* [64] demonstrated that a chevron-shaped boundary represents an excellent trapping device for self-propelled active particles. Accordingly, the deformation of the membrane induced by the active particles in the trapping state would eventually trap other particles, thus resulting into an increased penetration capability.

#### 4. Analytical theory

To rationalize our numerical results, we derive in the following an analytical theory based on a perturbative expansion technique that describes the system behavior in the small-deformation regime considering one active particle. Particularly, we are interested to determine theoretically the membrane displacement field of the particles in the trapping state. Our analytical calculations proceed through the linearization of the governing equations of motion, followed by prescribing the relevant fields using a discrete-to-continuum approach [105, 106].

#### 4.1. Linearized equations of motion

In the following, we neglect for simplicity the steric interactions between the membrane particles and assume that the mutual distance between neighboring particle is within the interaction range of the elastic forces, i.e.  $r_{ij} \in [h, \ell]$ , with  $j \in \mathcal{N}(i)$ , for i = 1, ..., N.

The dynamical equation governing the evolution of the *i*th membrane particle displacement field can be cast in the form

$$\dot{r}_i = \mu(F_i^{\rm E} + F_i^{\rm ext}),\tag{9}$$

wherein the superposed dot represents a temporal derivative, and  $F_i^E$  is the elastic force.



**Figure 4.** State diagram in the parameter spaces (a)  $(\delta, E)$  for  $\lambda = 3/2$  and (b)  $(\delta, \lambda)$  for E = 1, where, in both diagrams,  $\kappa = 10^{-2}$ . Symbols represent the dynamical state resulting from numerical integration of the governing equations of motion given by equations (2) and (3). In addition to trapping (blue squares) and penetration with healing (red triangles), penetration events without subsequent healing (green symbols) occur in some parameter ranges for large values of the size ratio  $\delta$  and reduced activity *E* for  $\lambda \leq 3/2$ . These penetration scenarios may be accompanied by the creation of a permanent hole of a size exceeding the interaction range of the membrane particles in addition to the partial fragmentation of the membrane into isolated or clusters of particles.

**Table 2.** Initial Cartesian coordinates of the  $N_{\rm P}$  active particles in the transverse plane.

$\overline{N_{\mathrm{P}}}$	Scaled initial positions $(x, y)/(h\sqrt{2})$
1	(0,0)
4	$(\pm 1/2, \pm 1/2)$
8	$(\pm 1/2, \pm 1/2), (0, \pm 1), (\pm 1, 0)$
12	$(\pm 1/2, \pm 1/2), (0, \pm 1), (\pm 1, 0), (\pm 1, \pm 1)$
24	$(\pm 1/2, \pm 1/2), (0, \pm 1), (\pm 1, 0), (\pm 1, \pm 1)$
	$(\pm 2, 0), (0, \pm 2), (\pm 3/2, \pm 1/2), (\pm 1/2, \pm 3/2)$





**Figure 6.** State diagram in the parameter space ( $\kappa$ ,  $N_P$ ) for  $\delta = 1$ ,  $\lambda = 3/2$ , and E = 1. Symbols represent the state resulting from numerical integration of the governing equations of motion given by equations (2) and (3) for  $N_P$  active particles initially placed on a square lattice around the center of the membrane. The parameter *r* quantifies the fraction of the active particles that penetrate through the membrane.

Assuming that the active particle has a radius comparable to that of the membrane particles, i.e. for  $\delta \sim 1$ , it can readily be verified that the resistive force due to the steric interactions with the active particle vanishes except for the four particles located near the center of the membrane, the initial coordinates of which are given in the Cartesian coordinate system by  $(x, y) = (\pm \sqrt{2} h/2, 0)$  and  $(x, y) = (0, \pm \sqrt{2} h/2)$ .

Following a linear elasticity theory approach [107, 108], we express the position vectors of each particle relative to the laboratory frame as  $r_i = (U_i + u_i)\hat{e}_x + (V_i + v_i)\hat{e}_y + w_i\hat{e}_z$ , for i = 1, ..., N, where  $U_i\hat{e}_x + V_i\hat{e}_y$  is the position vector in the undeformed state of reference, and  $u_i\hat{e}_x + v_i\hat{e}_y + w_i\hat{e}_z$  is the displacement of the membrane particles relative to the initial configuration. The linearized elastic force acting on the *i*th particle reads

$$F_{i}^{\rm E} = -\frac{kh}{2} \begin{cases} 2(p_{1}+p_{5}) + (2-\xi)S_{p} + 2(1-\xi)(p_{3}+p_{7}) + \xi(q_{2}-q_{4}+q_{6}-q_{8})\\ \xi(p_{2}-p_{4}+p_{6}-p_{8}) + 2(1-\xi)(q_{1}+q_{5}) + (2-\xi)S_{q} + 2(q_{3}+q_{7})\\ 2(1-\xi)(S_{r}+Q_{r}) \end{cases} \end{cases}$$
(10)

where we have defined  $p_j = (u_i - u_j)/h$ ,  $q_j = (v_i - v_j)/h$ , and  $r_j = (w_i - w_j)/h$  to denote the displacement gradients. Here, the numbers j = 1, ..., 8 appearing in subscript denote the index of a nearest or next-nearest-neighbor particle on the lattice, as schematically illustrated in figure 1(b). Moreover, we have used the shorthand notations  $S_{\alpha} = \alpha_2 + \alpha_4 + \alpha_6 + \alpha_8$  and  $Q_{\alpha} = \alpha_1 + \alpha_3 + \alpha_5 + \alpha_7$ , for  $\alpha \in \{p, q, r\}$ .

Notably, the inplane components of the elastic forces involve gradients of the lateral displacements  $p_j$  and  $q_j$ . In contrast to that, the normal components are found to depend on the displacement gradient  $r_j$  only. Consequently, a decoupling between the lateral and normal displacements is found for planar membranes, in a way analogous to what has previously been observed for 2D elastic membranes that are modeled as a continuum hyperelastic material featuring resistance toward shear and bending [109–113]. Particularly, for a nonprestressed membrane ( $\xi = 1$ ), the elastic forces are purely tangential (oriented along the plane of the membrane) and depend solely on the inplane displacement gradients  $p_j$  and  $q_j$ .

Having derived linearized expressions for the forces and torques governing the evolution of the membrane particles, we next consider the dynamics of the active particle. The latter is subject to the active driving force  $F_0 = F_0 \hat{e}_z$  in addition to the repulsive steric forces resulting from the interaction with the nearby membrane particles. In the overdamped regime, the translational motion of the active particle along the *z* direction is governed by

$$6\pi\eta R \,\dot{z}_{\rm P} = F_0 - 4F^{\rm ext}\cos\alpha,\tag{11}$$

wherein  $z_P$  denotes the z-position of the active particle,  $F^{\text{ext}}$  stands for the magnitude of the steric force exerted by one of the four particles located around the membrane center, and  $\alpha$  denotes the angle this force makes with the vertical.

Equations (9) form *N* ordinary differential equations in the time variable for the unknown membrane displacement field. These equations are subject to the initial conditions of vanishing membrane displacement, in addition to vanishing displacement at the membrane periphery and periodic boundary conditions along the *x* and *y* directions. In the steady state, the problem is equivalent to searching for the solution of linear recurrence relations coupling the positions of all the membrane particles initially located on a lattice. Due to the somewhat complicated nature of the resulting equations, an analytical solution is far from being trivial. To handle this difficulty and to obtain a quantitative insight into the system behavior in the small-deformation regime, we will approach the problem differently. Our solution methodology will be based on a continuum description of the linearized equations of motion as detailed below.

#### 4.2. Continuum theory

The core idea of discrete-to-continuum analysis is to express the membrane displacements following the standard approach as

$$\begin{pmatrix} u_{i+s,i+r} \\ v_{i+s,i+r} \\ w_{i+s,i+r} \end{pmatrix} = \exp(h\sqrt{2}(sD_x + rD_y)) \begin{pmatrix} u(x, y) \\ v(x, y) \\ w(x, y) \end{pmatrix},$$
(12)

where  $D_{\alpha} = \partial/\partial \alpha$ ,  $\alpha \in \{x, y\}$  represents the differential operator and  $(s, r) \in \{0, \pm 1/2, \pm 1\}$ . Here, the fractions at subscripts  $i \pm 1/2$  refer to the nearest-neighboring particle on the lattice axes, namely, the ones identified by even numbers in figure 1(b). The integer subscripts  $i \pm 1$  refer to the next-nearest-neighboring particles located on the lattice diagonals.

The exponential argument in equation (12) can be expanded up to the second order in power series using a two-dimensional Taylor expansion as [114]

$$\exp(h\sqrt{2}(sD_x + rD_y)) = 1 + h\sqrt{2}(sD_x + rD_y) + h^2(s^2D_x^2 + 2srD_xD_y + r^2D_y^2) + \cdots.$$
(13)

Applying this transformation rule to equation (9), the partial differential equation governing the translational degrees of freedom of the membrane particles can be rewritten in vector form as

$$\begin{pmatrix} u_{,t} \\ v_{,t} \\ w_{,t} \end{pmatrix} = \frac{A}{2} \begin{pmatrix} (6-\xi)u_{,xx} + (6-5\xi)u_{,yy} + 2\xi v_{,xy} \\ (6-5\xi)v_{,xx} + (6-\xi)v_{,yy} + 2\xi u_{,xy} \\ 6(1-\xi)(w_{,xx} + w_{,yy}) \end{pmatrix} + h^2(\mu F_0 - \delta \dot{z}_{\rm P})\delta(x,y)\hat{e}_z,$$
(14)

where  $A := \mu kh^2$  is a parameter having the dimension of a diffusion coefficient. Here, we have approximated the steric force exerted on the particles near the center of the membrane by a two-dimensional Dirac delta function  $\delta(x, y) = \delta(x)\delta(y)$ .

We have checked that taking alternative forms for the steric force, such as a 2D rectangle function centered around the origin, does not alter our results significantly. Therefore, a Dirac delta function has been adopted here for simplicity.

In the steady-state limit, equation (14) simplifies to

$$\begin{pmatrix} (6-\xi)u_{,xx} + (6-7\xi)u_{,yy} \\ (6-7\xi)v_{,xx} + (6-\xi)v_{,yy} \\ 6(1-\xi)(w_{,xx} + w_{,yy}) \end{pmatrix} + hP_0\delta(x,y)\,\hat{\boldsymbol{e}}_z = \boldsymbol{0},$$
(15)

wherein  $P_0 = 2F_0/(kh)$ , is the system admittance defined above by equation (8). In equation (15), we explicitly observe that  $P_0$  controls how far the active particle can penetrate into the initial planar membrane and lead to a deflection of the membrane. We note that the 2D Dirac delta function has the dimension of inverse length squared. In the following, we attempt to obtain closed analytical expressions for the displacement field not only for the steady state but also for transient dynamics situations.

#### 4.3. Steady solution

Because of the already-mentioned decoupling between the lateral and normal displacements, the solution for the in- and out-of-plane deformations can be obtained independently. Since the external force is exerted normal to the plane of the membrane, deformation will predominantly occur along the *z* direction. In the following, we assume that  $|w| \ll L$ , for our approximate equations of motion derived above to be valid.

By projecting equation (15) onto the *z* direction, the normal displacement is governed by a second-order partial differential equation of the form

$$6(1 - \xi)(w_{,xx} + w_{,yy}) + hP_0 \,\delta(x, y) = 0.$$
<sup>(16)</sup>

To solve equation (16), we exploit periodicity of the system along the transverse directions by expressing the membrane normal displacement *w* in terms of a Fourier series [115]. Then,

$$w(x, y) = \frac{4}{L^2} \sum_{p \ge 1} \sum_{q \ge 1} \hat{w}(p, q) c_p(x) c_q(y),$$
(17)

with p, q = 1, 2, ... denoting the positive integers that set the coordinates in Fourier space. Here, we have defined the basis function  $c_p(x) = \cos(H_p x)$ , where  $H_p = (2p - 1)\pi/L$ , and analogously for  $c_q(y)$ . In addition,  $\hat{w}(p, q)$  denotes the Fourier coefficients of w, defined as

$$\hat{w}(p,q) = \int_{-\frac{L}{2}}^{\frac{L}{2}} \int_{-\frac{L}{2}}^{\frac{L}{2}} w(x,y) c_p(x) c_q(y) \, \mathrm{d}x \, \mathrm{d}y.$$
(18)

It is worth mentioning that the solution form given by equation (17) follows from the prescribed boundary conditions, so as to ensure that  $w(x = \pm L/2, y) = w(x, y = \pm L/2) = 0$ . Moreover, the basis functions  $c_p(x)$  satisfy the orthogonality relation

$$\int_{-\frac{L}{2}}^{\frac{L}{2}} c_p(x) c_{p'}(x) \, \mathrm{d}x = \frac{L}{2} \, \delta_{pp'}.$$
(19)

By substituting equation (17) into (16) and making use of the orthogonality property given by equation (19), we readily obtain

$$\hat{w}(p,q) = \frac{hP_0}{6(1-\xi)(H_p^2 + H_q^2)}.$$
(20)

Finally, by writing the solution for the transverse displacements u(x, y) and v(x, y) in terms of Fourier series in a way analogous to equation (17), it follows that u and v must vanish to satisfy the boundary conditions imposed at the membrane extremities, considering the present approximate equations.

Figure 7 shows the steady-state variations of the normal displacement (scaled by the membrane size) versus x/L. Results are shown in the plane of maximum deformation y = 0 for three different values of the reduced activity *E*, while keeping the reduced elasticity to  $\kappa = 1$ . Symbols indicate the numerical solution of the full nonlinear problem given by equations (2) and (3) and solid lines are the analytical predictions obtained from the solution of the continuum equations using finite Fourier transforms. Good agreement is found between the theory and simulations. All in all, our predictive model requires no fitting parameters and thus can conveniently be applied to describe the steady-state membrane displacement in the small-deformation regime considered here.

#### 4.4. Transient dynamics

Having investigated the system behavior in the steady trapping limit, we next turn our attention to the transient dynamics under the action of the force exerted by an active particle pushing against the membrane. To be able to







**Figure 8.** Membrane normal displacement out of the plane of the undeformed membrane as functions of scaled distance x/L calculated in the plane y = 0. Comparison is made between analytical predictions (lines) and full numerical simulations (symbols) for the transient behavior before the trapping state at various scaled times where, again,  $t_S = \eta L^3/\epsilon$  denotes the unit of simulation time. Here, we set ( $\kappa, E$ ) = (1, 1).

make an analytical progress, we assume that  $\delta \dot{z}_P \ll \mu F_0$ , such that  $F_0$  is balanced by the steric interaction with the membrane, not by friction with the fluid. Accordingly, we set  $\dot{z}_P = 0$  in equation (14) for t > 0.

The projected equation of motion governing the temporal evolution of the normal displacement field *w* reads

$$\frac{w_{,t}}{A} = 3(1-\xi)(w_{,xx}+w_{,yy}) + \frac{hP_0}{2}\,\delta(x,y).$$
(21)

Using a similar solution procedure as for the steady dynamics that is based on Fourier transforms, we obtain

$$\frac{\hat{w}_{,t}}{A} = -3(1-\xi)(H_p^2 + H_q^2)\hat{w} + \frac{hP_0}{2}.$$
(22)

Applying Laplace transforms [116] to equation (22) and solving for the unknown field  $\hat{w}$ , we readily obtain

$$\hat{w}(p,q) = \frac{hP_0A}{2s(3A(1-\xi)(H_p^2 + H_q^2) + s)}.$$
(23)

The expression of the Fourier coefficient in the time domain follows for thwith by inverse Laplace transform as

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$$\frac{\hat{w}}{\hat{w}_{\infty}} = 1 - e^{-3A(1-\xi)(H_p^2 + H_q^2)t},$$

where  $\hat{w}_{\infty}$  represents the steady normal displacement given by equation (20).

In figure 8, we present the transient evolution of the membrane normal displacement before reaching the steady state at three scaled times, where  $t_S = \eta L^3/\epsilon$  denotes the simulation time. Here, curves are shown in the plane y = 0 using the membrane parameters ( $\kappa, E$ ) = (1, 1). As time increases, the membrane deformation exponentially approaches the steady-state value. Although the analytical theory involves no fitting parameters, very good agreement is obtained between full numerical simulations (symbols) and analytical predictions (lines).

# 5. Conclusion

In the present work, we have discussed the interaction of an active particle with a minimal 2D membrane which could be realized, e.g using synthetic particles of controlled interactions. We have identified three different scenarios, one corresponding to a permanent trapping of the particle by the membrane and the remaining two implying penetration of the particle through the membrane. The first type of penetration is characterized by a complete subsequent healing of the membrane which relaxes towards its equilibrium configuration once the particle has passed. In stark contrast, we have shown that much larger particles can create a hole in the membrane that is large enough to prevent such a self-healing dynamics, resulting in a permanently damaged membrane. This behavior is accompanied by the expulsion of membrane particles into isolated fragments. Our result suggests that if one were to effectively damage a synthetic vesicle, or perhaps a cancer cell membrane, one would need to use particles of a certain minimal size. Complementary to simulations, we here provide a detailed analytical theory allowing to predict the entire state diagram, the shape and the dynamics of the membrane. Our approach might be useful to predict transitions between trapping, penetration with and without self-healing in experiments.

# Acknowledgments

We thank Joachim Clement and Cornelia Monzel for stimulating discussions. The authors gratefully acknowledge support from the DFG (Deutsche Forschungsgemeinschaft) through the projects DA 2107/1-1, ME 3571/2-2, and LO 418/16-3.

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## References

- [1] Budin I and Devaraj N K 2011 Membrane assembly driven by a biomimetic coupling reaction J. Am. Chem. Soc. 134751-3
- [2] Osada Y and Nakagawa T 1992 Membrane Science and Technology (Boca Raton, FL: CRC Press) (https://doi.org/10.1002/ pi.1994.210330425)
- [3] Belfort G 2012 Synthetic Membrane Process: Fundamentals and Water Applications (Orlando, FL: Academic) (https://doi.org/ 10.1002/aic.690329821)
- [4] Brown A T and Cicuta P 2017 Biological fluid interfaces and membranes The Oxford Handbook of Soft Condensed Matter (Oxford: Oxford University Press) (https://doi.org/10.1093/oxfordhb/9780199667925.001.0001)
- [5] Fane A G, Wang R and Hu M X 2015 Synthetic membranes for water purification: status and future Angew. Chem. Int. Ed. 54 3368
- [6] Kocsis I et al 2018 Oriented chiral water wires in artificial transmembrane channels Sci. Adv. 4 eaao5603
- [7] Vienken J, Diamantoglou M, Henne W and Nederlof B 1999 Artificial dialysis membranes: from concept to large scale production *Am. J. Nephrol.* 19 355–62
- [8] Klinkmann H and Vienken J 1995 Membranes for dialysis Nephrol. Dial. Transplant. 10 39
- [9] Davis A P, Sheppard D N and Smith B D 2007 Development of synthetic membrane transporters for anions Chem. Soc. Rev. 36 348-57
- [10] Kim Y-R, Jung S, Ryu H, Yoo Y-E, Kim S M and Jeon T-J 2012 Synthetic biomimetic membranes and their sensor applications Sensors 12 9530–50
- [11] Jun I-K and Hess H 2010 A biomimetic, self-pumping membrane Adv. Mater. 22 4823-5
- [12] Langer R 1998 Drug delivery and targeting Nature 392 5-10
- [13] Verma A, Uzun O, Hu Y, Hu Y, Han H-S, Watson N, Chen S, Irvine D J and Stellacci F 2008 Surface-structure-regulated cellmembrane penetration by monolayer-protected nanoparticles Nat. Mater. 7 588–95
- [14] Yang K and Ma Y-Q 2010 Computer simulation of the translocation of nanoparticles with different shapes across a lipid bilayer Nat. Nanotechnol. 5 579
- [15] Lin J, Zhang H, Chen Z and Zheng Y 2010 Penetration of lipid membranes by gold nanoparticles: insights into cellular uptake, cytotoxicity, and their relationship ACS Nano 4 5421–9

- [16] Verma A and Stellacci F 2010 Effect of surface properties on nanoparticle-cell interactions Small 6 12-21
- [17] Nel A E, M\u00e4dler L, Velegol D, Xia T, Hoek E M, Somasundaran P, Klaessig F, Castranova V and Thompson M 2009 Understanding biophysicochemical interactions at the nano-bio interface Nat. Mater. 8 543
- [18] Wang T, Bai J, Jiang X and Nienhaus G U 2012 Cellular uptake of nanoparticles by membrane penetration: a study combining confocal microscopy with FTIR spectroelectrochemistry ACS Nano 6 1251–9
- [19] Shang L, Nienhaus K and Nienhaus G U 2014 Engineered nanoparticles interacting with cells: size matters J. Nanobiotechnol. 125
- [20] Monzel C, Vicario C, Piehler J, Coppey M and Dahan M 2017 Magnetic control of cellular processes using biofunctional nanoparticles Chem. Sci. 8 7330–8
- [21] Liße D, Monzel C, Vicario C, Manzi J, Maurin I, Coppey M, Piehler J and Dahan M 2017 Engineered ferritin for magnetogenetic manipulation of proteins and organelles inside living cells Adv. Mater. 29 1700189
- [22] Müller E K, Gräfe C, Wiekhorst F, Bergemann C, Weidner A, Dutz S and Clement J H 2018 Magnetic nanoparticles interact and pass an *in vitro* co-culture blood-placenta barrier model *Nanomaterials* **8** 108
- [23] Xuan M, Shao J, Gao C, Wang W, Dai L and He Q 2018 Self-propelled nanomotors for thermomechanically percolating cell membranes Angew. Chem. 130 12643–7
- [24] Wang W, Wu Z, Lin X, Si T and He Q 2019 Gold nanoshell-functionalized polymer nanoswimmer for photomechanical poration of single cell membrane J. Am. Chem. Soc. 141 6601–8
- [25] Chithrani B D, Ghazani A A and Chan W C W 2006 Determining the size and shape dependence of gold nanoparticle uptake into mammalian cells Nano Lett. 6 662–8
- [26] Yang K and Ma Y Q 2010 Computer simulation of the translocation of nanoparticles with different shapes across a lipid bilayer Nat. Nanotechnol. 5 579–83
- [27] Dos Santos T, Varela J, Lynch I, Salvati A and Dawson K A 2011 Quantitative assessment of the comparative nanoparticle-uptake efficiency of a range of cell lines Small 7 3341–9
- [28] Parodi A et al 2013 Synthetic nanoparticles functionalized with biomimetic leukocyte membranes possess cell-like functions Nat. Nanotechnol. 8 61
- [29] Daddi-Moussa-Ider A, Guckenberger A and Gekle S 2016 Long-lived anomalous thermal diffusion induced by elastic cell membranes on nearby particles *Phys. Rev.* E 93 012612
- [30] Jünger F, Kohler F, Meinel A, Meyer T, Nitschke R, Erhard B and Rohrbach A 2015 Measuring local viscosities near plasma membranes of living cells with photonic force microscopy *Biophys. J.* 109 869–82
- [31] Gräfe C, Slabu I, Wiekhorst F, Bergemann C, von Eggeling F, Hochhaus A, Trahms L and Clement J 2016 Magnetic particle spectroscopy allows precise quantification of nanoparticles after passage through human brain microvascular endothelial cells *Phys. Med. Biol.* 61 3986
- [32] Ramaswamy S and Rao M 2001 The physics of active membranes C. R. Acad. Sci. IV 2817-39
- [33] Yoon Y Z, Kotar J, Brown A T and Cicuta P 2011 Red blood cell dynamics: from spontaneous fluctuations to non-linear response Soft Matter 7 2042–51
- [34] Lacoste D and Bassereau P 2014 An update on active membrane *Lipid Bilayers and Model Membranes: From Basic Research to Application* vol 271 (Boca Raton, FL: CRC Press) (https://doi.org/10.1201/b16617)
- [35] Mitragotri S and Lahann J 2009 Physical approaches to biomaterial design Nat. Mater. 8 15
- [36] Li Y, Li X, Li Z and Gao H 2012 Surface-structure-regulated penetration of nanoparticles across a cell membrane *Nanoscale* 4 3768–75
- [37] Wong-Ekkabut J, Baoukina S, Triampo W, Tang I-M, Tieleman D P and Monticelli L 2008 Computer simulation study of fullerene translocation through lipid membranes Nat. Nanotechnol. 3 363
- [38] Junot G, Briand G, Ledesma-Alonso R and Dauchot O 2017 Active versus passive hard disks against a membrane: mechanical pressure and instability Phys. Rev. Lett. 119 028002
- [39] Marconi U M B, Sarracino A, Maggi C and Puglisi A 2017 Self-propulsion against a moving membrane: enhanced accumulation and drag force Phys. Rev. E 96 032601
- [40] Costanzo A, Elgeti J, Auth T, Gompper G and Ripoll M 2014 Motility-sorting of self-propelled particles in microchannels EPL 107 36003
- [41] Daddi-Moussa-Ider A, Goh S, Liebchen B, Hoell C, Mathijssen A J, Guzmán-Lastra F, Scholz C, Menzel A M and Löwen H 2019 Membrane penetration and trapping of an active particle J. Chem. Phys. 150 064906
- [42] Boal D H, Seifert U and Zilker A 1992 Dual network model for red blood cell membranes *Phys. Rev. Lett.* 69 3405
- [43] Lidmar J, Mirny L and Nelson D R 2003 Virus shapes and buckling transitions in spherical shells Phys. Rev. E 68 051910
- [44] Noguchi H and Gompper G 2005 Shape transitions of fluid vesicles and red blood cells in capillary flows Proc. Natl Sci. USA 102 14159
- [45] Fedosov D A, Caswell B and Karniadakis G E 2010 Systematic coarse-graining of spectrin-level red blood cell models Comput. Methods Appl. Mech. Eng. 199 1937–48
- [46] Förtsch A, Laumann M, Kienle D and Zimmermann W 2017 Migration reversal of soft particles in vertical flows Europhys. Lett. 119 64003
- [47] Laumann M, Bauknecht P, Gekle S, Kienle D and Zimmermann W 2017 Cross-stream migration of asymmetric particles driven by oscillating shear Europhys. Lett. 117 44001
- [48] Laumann M, Schmidt W, Farutin A, Kienle D, Förster S, Misbah C and Zimmermann W 2019 Emerging attractor in wavy poiseuille flows triggers sorting of biological cells *Phys. Rev. Lett.* 122 128002
- [49] Froltsov V, Blaak R, Likos C and Löwen H 2003 Crystal structures of two-dimensional magnetic colloids in tilted external magnetic fields Phys. Rev. E 68 061406
- [50] Barry E and Dogic Z 2010 Entropy driven self-assembly of nonamphiphilic colloidal membranes Proc. Natl Acad. Sci. USA 107 10348–53
- [51] Ewerlin M, Demirbas D, Brüssing F, Petracic O, Ünal A A, Valencia S, Kronast F and Zabel H 2013 Magnetic dipole and higher pole interaction on a square lattice Phys. Rev. Lett. 110 177209
- [52] Messina R, Khalil L A and Stanković I 2014 Self-assembly of magnetic balls: from chains to tubes Phys. Rev. E 89 011202
- [53] Kaiser A, Popowa K and Löwen H 2015 Active dipole clusters: from helical motion to fission *Phys. Rev. E* 92 012301
- [54] Guzmán-Lastra F, Kaiser A and Löwen H 2016 Fission and fusion scenarios for magnetic microswimmer clusters Nat. Commun. 7 13519
- [55] Yener A B and Klapp S H L 2016 Self-assembly of three-dimensional ensembles of magnetic particles with laterally shifted dipoles Soft Matter 12 2066–75
- [56] Spiteri L and Messina R 2017 Columnar aggregation of dipolar chains Europhys. Lett. 120 36001
- [57] Messina R and Stanković I 2017 Assembly of magnetic spheres in strong homogeneous magnetic field Physica A 466 10-20

- [58] Peroukidis S D and Klapp S H L 2016 Orientational order and translational dynamics of magnetic particle assemblies in liquid crystals Soft Matter 12 6841–50
- [59] Deißenbeck F, Löwen H and Oğuz E C 2018 Ground state of dipolar hard spheres confined in channels Phys. Rev. E 97 052608
- [60] García-Torres J, Calero C, Sagués F, Pagonabarraga I and Tierno P 2018 Magnetically tunable bidirectional locomotion of a selfassembled nanorod-sphere propeller Nat. Commun. 9 1663

[61] Oğuz E C, Mijailović A and Schmiedeberg M 2018 Self-assembly of complex structures in colloid-polymer mixtures Phys. Rev. E 98 052601

- [62] ten Hagen B, van Teeffelen S and Löwen H 2011 Brownian motion of a self-propelled particle J. Phys.: Condens. Matter 23 194119
- [63] Wittkowski R and Löwen H 2012 Self-propelled brownian spinning top: dynamics of a biaxial swimmer at low reynolds numbers Phys. Rev. E 85 021406
- [64] Kaiser A, Wensink H and Löwen H 2012 How to capture active particles Phys. Rev. Lett. 108 268307
- [65] Wensink H H and Löwen H 2012 Emergent states in dense systems of active rods: from swarming to turbulence J. Phys.: Condens. Matter 24 464130
- [66] Kümmel F, ten Hagen B, Wittkowski R, Buttinoni I, Eichhorn R, Volpe G, Löwen H and Bechinger C 2013 Circular motion of asymmetric self-propelling particles Phys. Rev. Lett. 110 198302
- [67] Ten Hagen B, Kümmel F, Wittkowski R, Takagi D, Löwen H and Bechinger C 2014 Gravitaxis of asymmetric self-propelled colloidal particles Nat. Commun. 5 4829
- [68] ten Hagen B, Wittkowski R, Takagi D, Kümmel F, Bechinger C and Löwen H 2015 Can the self-propulsion of anisotropic microswimmers be described by using forces and torques? J. Phys. Condes. Matter 27 194110
- [69] Speck T and Jack R L 2016 Ideal bulk pressure of active Brownian particles Phys. Rev. E 93 062605
- [70] de Graaf J, Menke H, Mathijssen A J T M, Fabritius M, Holm C and Shendruk T N 2016 Lattice-Boltzmann hydrodynamics of anisotropic active matter J. Chem. Phys. 144 134106
- [71] Liebchen B, Marenduzzo D and Cates M E 2017 Phoretic interactions generically induce dynamic clusters and wave patterns in active colloids Phys. Rev. Lett. 118 268001
- [72] Hoell C, Löwen H and Menzel A M 2017 Dynamical density functional theory for circle swimmers New J. Phys. 19 125004
- [73] Daddi-Moussa-Ider A and Menzel A M 2018 Dynamics of a simple model microswimmer in an anisotropic fluid: implications for alignment behavior and active transport in a nematic liquid crystal Phys. Rev. Fluids 3 094102
- [74] Lauga E and Powers T R 2009 The hydrodynamics of swimming microorganisms Rep. Prog. Phys. 72 096601
- [75] Zöttl A and Stark H 2016 Emergent behavior in active colloids J. Phys.: Condens. Matter 28 253001
- [76] Lauga E 2016 Bacterial hydrodynamics Ann. Rev. Fluid Mech. 48 105–30
- [77] Elgeti J, Winkler R G and Gompper G 2015 Physics of microswimmers- single particle motion and collective behavior: a review Rep. Prog. Phys. 78 056601
- [78] Bechinger C, Di Leonardo R, Löwen H, Reichhardt C, Volpe G and Volpe G 2016 Active particles in complex and crowded environments Rev. Mod. Phys. 88 045006
- [79] Illien P, Golestanian R and Sen A 2017 Fuelled motion: phoretic motility and collective behaviour of active colloids Chem. Soc. Rev. 46 5508–18
- [80] Gao W, Feng X, Pei A, Kane C R, Tam R, Hennessy C and Wang J 2013 Bioinspired helical microswimmers based on vascular plants Nano Lett. 14 305–10
- [81] Gao W and Wang J 2014 Synthetic micro/nanomotors in drug delivery Nanoscale 6 10486-94
- [82] Scholz C, Engel M and Pöschel T 2018 Rotating robots move collectively and self-organize Nat. Commun. 9 931
- [83] Scholz C, Jahanshahi S, Ldov A and Löwen H 2018 Inertial delay of self-propelled particles *Nat. Commun.* **9** 5156
- [84] Adey WR 1977 Models of membranes of cerebral cells as substrates for information storage Biosystems 8 163-78
- [85] Jiang J, Eisenthal K B and Yuste R 2007 Second harmonic generation in neurons: electro-optic mechanism of membrane potential sensitivity *Biophys. J.* 93 L26–8
- [86] Demirörs A F, Stiefelhagen J C, Vissers T, Smallenburg F, Dijkstra M, Imhof A and van Blaaderen A 2015 Long-ranged oppositely charged interactions for designing new types of colloidal clusters *Phys. Rev.* X 5 021012
- [87] Gov N S 2007 Active elastic network: cytoskeleton of the red blood cell Phys. Rev. E 75 011921
- [88] Pivkin I V and Karniadakis G E 2008 Accurate coarse-grained modeling of red blood cells Phys. Rev. Lett. 101 118105
- [89] Peng Z, Li X, Pivkin I V, Dao M, Karniadakis G E and Suresh S 2013 Lipid bilayer and cytoskeletal interactions in a red blood cell Proc. Natl Acad. Sci. USA 110 13356–61
- [90] Mejean C O, Schaefer A W, Buck K B, Kress H, Shundrovsky A, Merrill J W, Dufresne E R and Forscher P 2013 Elastic coupling of nascent apcam adhesions to flowing actin networks PLoS One 8 e73389
- [91] Weeks J D, Chandler D and Andersen H C 1971 Role of repulsive forces in determining the equilibrium structure of simple liquids J. Chem. Phys. 54 5237–47
- [92] Babel S, Löwen H and Menzel A M 2016 Dynamics of a linear magnetic microswimmer molecule Europhys. Lett. 113 58003
- [93] Happel J and Brenner H 2012 Low Reynolds Number Hydrodynamics: With Special Applications to Particulate Media (Dordrecht: Springer) (https://doi.org/10.1007/978-94-009-8352-6)
- [94] Kim S and Karrila S J 2013 Microhydrodynamics: Principles and Selected Applications (Mineola, NY: Courier Corporation)
- [95] Swan J W and Brady J F 2007 Simulation of hydrodynamically interacting particles near a no-slip boundary Phys. Fluids 19 113306
- [96] Swan J W and Brady J F 2010 Particle motion between parallel walls: hydrodynamics and simulation *Phys. Fluids* 22 103301
- [97] Balboa-Usabiaga F, Kallemov B, Delmotte B, Bhalla A, Griffith B and Donev A 2017 Hydrodynamics of suspensions of passive and active rigid particles: a rigid multiblob approach Commun. Appl. Math. Comput. Sci. 11 217–96
- [98] Driscoll M and Delmotte B 2018 Leveraging collective effects in externally driven colloidal suspensions: experiments and simulations Curr. Opin. Colloid Interface Sci. 40 42–57
- [99] Press W H 1992 The Art of Scientific Computing (Cambridge: Cambridge University Press)
- [100] Jiang W, Kim B Y, Rutka J T and Chan W C 2008 Nanoparticle-mediated cellular response is size-dependent *Nat. Nanotechnol.* 3 145
   [101] Andersson P O, Lejon C, Ekstrand-Hammarström B, Akfur C, Ahlinder L, Bucht A and Österlund L 2011 Polymorph-and size-
- dependent uptake and toxicity of tio2 nanoparticles in living lung epithelial cells Small 7 514–23
   [102] Oh E et al 2011 Cellular uptake and fate of pegylated gold nanoparticles is dependent on both cell-penetration peptides and particle size ACS Nano 5 6434–48
- [103] Huang W, Chang C B and Sung H J 2012 Three-dimensional simulation of elastic capsules in shear flow by the penalty immersed boundary method J. Comput. Phys. 231 3340–64

- [104] Elbakry A, Wurster E-C, Zaky A, Liebl R, Schindler E, Bauer-Kreisel P, Blunk T, Rachel R, Goepferich A and Breunig M 2012 Layerby-layer coated gold nanoparticles: Size-dependent delivery of DNA into cells Small 8 3847–56
- [105] Goh S, Menzel A M and Löwen H 2018 Dynamics in a one-dimensional ferrogel model: relaxation, pairing, shock-wave propagation Phys. Chem. Chem. Phys. 20 15037–51
- [106] Menzel A M 2019 Mesoscopic characterization of magnetoelastic hybrid materials: magnetic gels and elastomers, their particle-scale description, and scale-bridging links Arch. Appl. Mech. 89 17–45
- [107] Timoshenko S P and Woinowsky-Krieger S 1959 Theory of Plates and Shells vol 2 (New York: McGraw-Hill)
- [108] Sadd MH 2009 Elasticity: Theory, Applications, and Numerics (Cambridge, MA: Academic)
- [109] Daddi-Moussa-Ider A, Guckenberger A and Gekle S 2016 Particle mobility between two planar elastic membranes: Brownian motion and membrane deformation Phys. Fluids 28 071903
- [110] Daddi-Moussa-Ider A, Rallabandi B, Gekle S and Stone H A 2018 Reciprocal theorem for the prediction of the normal force induced on a particle translating parallel to an elastic membrane *Phys. Rev. Fluids* 3 084101
- [111] Daddi-Moussa-Ider A and Gekle S 2018 Brownian motion near an elastic cell membrane: a theoretical study Eur. Phys. J. E 41 19
- [112] Daddi-Moussa-Ider A, Lisicki M, Gekle S, Menzel A M and Löwen H 2018 Hydrodynamic coupling and rotational mobilities near planar elastic membranes J. Chem. Phys. 149 014901
- [113] Daddi-Moussa-Ider A, Kaoui B and Löwen H 2019 Axisymmetric flow due to a stokeslet near a finite-sized elastic membrane J. Phys. Soc. Japan 88 054401
- [114] Rosenau P 2003 Hamiltonian dynamics of dense chains and lattices: or how to correct the continuum Phys. Lett. A 311 39–52
- [115] Bracewell R 1999 The Fourier Transform and its Applications (New York: McGraw-Hill)
- [116] Widder D V 2015 Laplace Transform (PMS-6) (Princeton, NJ: Princeton University Press)