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DNA repair synthesis and histone deposition partner during homologous recombination

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ABSTRACT

Chromatin remodeling is critical for the regulation of the DNA damage response. We highlight findings from our recent study showing that the deposition of the histone variant H3.3 by the alpha-thalassemia mental retardation X-linked protein (ATRX) and the death domain associated protein (DAXX) chromatin remodeling complex regulates DNA repair synthesis during homologous recombination.

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Chromatin modification and remodeling is a dynamic process that is established not only as prerequisite to allow access to DNA for repair following damage, but also as a key regulator of the DNA damage response (DDR). Chromatin dynamics during the DDR involve histone modifications, such as phosphorylation and methylation, and also the disassembly and reassembly of nucleosomes before and after DNA repair, respectively. The removal of original histones is often followed by the deposition of histone variants that play important roles in the stress response. Extensive efforts have been dedicated to identify the functions of histone variants in the DDR and the different repair mechanisms. Among the most critical lesions occurring in the cell are DNA double strand breaks (DSBs), which can be repaired by 2 major pathways: non-homologous end-joining (NHEJ) and homologous recombination (HR). HR can be regulated by the early deposition and removal of the histone variant H2A.Z, which has been shown to control resection, an important processing step on break ends that commits the cell to repair by HR.¹ Another variant, H3.3, has been shown to be incorporated rapidly after DNA damage and promotes NHEJ.² It is also required for post-repair chromatin assembly at sites of DSBs repaired by NHEJ.³ While it is clear that histone exchange can regulate early DSB repair steps, such as repair pathway choice, chromatin dynamics influencing later stages, particularly in HR, remain poorly characterized.

One distinction between NHEJ and HR is the requirement for a sister chromatid in HR which is used as a template for repair synthesis to allow the faithful restoration of the original sequence. Repair synthesis occurs in various repair mechanisms, such as nucleotide excision repair (NER), but its regulation during HR is not well understood in human cells.⁴ While it has been shown that repair synthesis in NER is concomitant with chromatin reconstitution through the chromatin

assembly factor-1 (CAF-1), whether a similar process occurs in HR has not been shown.⁵ In our recently published work, we describe a novel role for H3.3 deposition by the chromatin remodeler alpha-thalassemia mental retardation X-linked protein (ATRX) and the chaperon death domain associated protein (DAXX) in a sub-pathway of HR-mediated DSB repair.⁶ We show that cells lacking ATRX, DAXX or H3.3 have a defect in repairing ionizing radiation-induced DSBs by HR. Strikingly, cells lacking these factors exhibit a strong defect in extended DNA repair synthesis, indicating a role of the chromatin remodeling complex during this late HR step. We employed a system for the *in vivo* visualization of H3.3 dynamics and showed that newly synthesized H3.3 is incorporated at sites of DNA damage at both early and late time points. However, only late incorporation was ATRX dependent and was associated with DNA synthesis. To further consolidate the association of repair synthesis with nucleosome assembly, we identified a strong damage-induced interaction between ATRX and proliferating cell nuclear antigen, which is required for both repair synthesis and repair completion.

Our findings inspire a model in which the chromatin remodeler ATRX promotes extended DNA synthesis by simultaneously facilitating chromatin reconstitution. Unlike in NER, this is a prerequisite for repair completion as the disruption of this process impedes repair. One possible explanation for this discrepancy is the extent of repair synthesis, which is much longer during HR and possibly requires nucleosome reassembly to allow further progression of the DNA synthesis machinery. A tight coupling of histone deposition with DNA synthesis also occurs during replication, where the coordination of chromatin assembly and DNA synthesis is important for the regulation of elongation rate and fork progression.⁷ Additionally, histone deposition is important

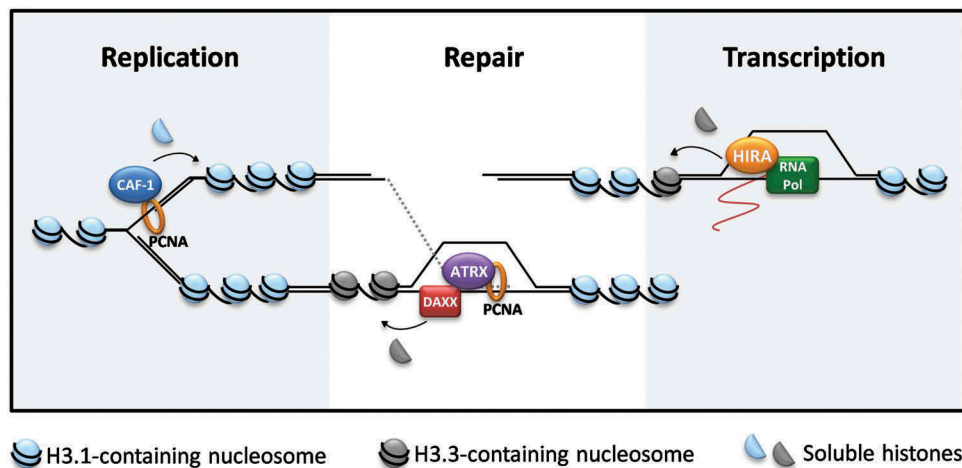


Figure 1. Histone dynamics in replication, repair synthesis and transcription. Chromatin remodeling is a common requirement for the regulation of DNA and RNA synthesis. Distinct chromatin remodelers and chaperons direct the deposition of histone variants during replication (chromatin assembly factor-1, CAF-1), repair synthesis in homologous recombination (alpha-thalassemia mental retardation X-linked protein, ATRX, and the chaperon death domain associated protein, DAXX) and transcription (histone regulator A, HIRA). PCNA: proliferating nuclear cell antigen; RNA Pol: RNA polymerase II.

during transcription and can influence transcription rate.⁸ Therefore, histone exchange seems to be critical for the regulation of processes that require extended DNA/RNA synthesis such as replication, transcription and repair (Figure 1). This is not surprising given the importance of maintaining genomic integrity in processes exposing naked DNA that needs protection, especially over long patches, a notion in agreement with the described gap-filling role of H3.3 during transcription.⁹ Furthermore, chromatin reconstitution can also serve a structural stabilization function to alleviate topological stress occurring during DNA unwinding, a major impediment for various DNA/RNA synthesis processes.

The role of ATRX during HR provides insight into how DNA repair synthesis is regulated *in vivo* and highlights the importance of histone variants and their remodelers in this process. It would be of interest to study similar processes in other repair pathways, which would establish a paradigm for chromatin reorganization during DNA repair synthesis. Additionally, this newly identified role for ATRX could help to better understand the diseases in which ATRX, H3.3 or DAXX are mutated. One prominent example is a subset of cancers that undergo a telomerase-independent telomere maintenance process known as the alternative lengthening of telomeres (ALT). ALT cancers represent about 10–15% of all cancers and almost always have ATRX, H3.3 or DAXX mutations.¹⁰ ALT is identified as an HR process that involves long range break-induced DNA synthesis. Given that ATRX's function is lost during ALT, it was surprising to uncover its HR role which raises the question of how this new role fits to the ALT process. One possibility is that ATRX has distinct functions at telomeres versus internal breaks. Alternatively, ATRX might promote a form of HR that is repressive to ALT and its loss allows HR that is more permissive to the ALT process. Delineating these functions is important for the understanding of ALT and can be of value for the development of therapeutic strategies for ALT cancers.

Disclosure of interest

The authors report no conflict of interest.

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